

Preview

Extracellular HSP90 warms up integrins for an irisin workout

Dimitra Bourboulia,^{1,2,3,*} Mark R. Woodford,^{1,2,3,*} and Mehdi Mollapour^{1,2,3,4,*}

¹Department of Urology, SUNY Upstate Medical University, Syracuse, NY 13210, USA

²Upstate Cancer Center, SUNY Upstate Medical University, Syracuse, NY 13210, USA

³Department of Biochemistry and Molecular Biology, SUNY Upstate Medical University, Syracuse, NY 13210, USA

⁴Twitter: @medmol

*Correspondence: bourmpod@upstate.edu (D.B.), woodform@upstate.edu (M.R.W.), mollapom@upstate.edu (M.M.)

<https://doi.org/10.1016/j.cmet.2023.06.002>

The hormone-like protein irisin is involved in browning of adipose tissue and regulation of metabolism. Recently, Mu et al. identified the extracellular chaperone heat shock protein-90 (Hsp90) as the activating factor for “opening” $\alpha V\beta 5$ integrin receptor, allowing for high-affinity irisin binding and effective signal transduction.

Is exercise good for you? The answer is obviously yes. Regular physical exercise has wide-ranging positive impacts on overall health and well-being, influencing various physiological systems and contributing to longevity and disease prevention. However, what are the immediate effects and consequences of exercise in the muscles? Previous work has shown that peroxisome proliferator-activated receptor- γ coactivator 1 alpha (PGC-1 α) plays a crucial role in regulating cellular energy metabolism and adaptive responses to physical exercise.¹ It is primarily expressed in tissues with high energy demands such as skeletal muscle, heart, liver, and brown adipose tissue. PGC-1 α is also responsible for the synthesis of plasma membrane-bound fibronectin type III domain-containing protein 5 (FNDC5).² The carboxy-terminal tail of FNDC5 is in the cytoplasm, whereas muscle contraction triggers proteolytic cleavage of an amino-terminal section of FNDC5 known as irisin. Irisin functions as a myokine, and studies in mice have shown that increasing irisin levels through genetic manipulation or exercise can lead to various health benefits including increased energy expenditure, improved insulin sensitivity, enhanced glucose uptake, and protection against diet-induced obesity.³ Additionally, irisin has been implicated in the regulation of bone health, cognitive function, and cardiovascular health. In bone, fat, and hippocampus, irisin tends to function primarily via αV integrin receptors, particularly $\alpha V\beta 5$.⁴ However, it is unclear how a small protein

like irisin interacts with and transduces signaling through an integrin receptor.

Mu et al.⁵ have now shown that irisin-mediated integrin activation uses a two-step process involving an extracellular molecular chaperone heat shock protein-90 α (eHsp90 α) (Figure 1). eHsp90 α is secreted from skeletal muscle and its secretion is elevated with exercise, although the total levels of cellular Hsp90 α in muscle remain unchanged. These findings together with the absence of other secreted chaperones suggested that eHsp90 α secretion is an exercise-induced mechanism and not simply released through cellular damage. The published work also indicated that eHsp90 α directly binds to the ectodomain of $\alpha V\beta 5$ and maintains it in an open conformation (Figure 1). Here eHsp90 α appears to function independent of ATP availability. This is an important consideration, as the extracellular ATP concentration is relatively low.⁶ The effect of Hsp90 α on irisin-mediated activation was tested in mice, and Hsp90 α neutralizing antibody treatment caused a significant reduction in irisin-induced integrin signaling. The $\alpha V\beta 5$ receptor, once activated by eHsp90 α , has a very high affinity (Kd \sim 30 nM) for the highly glycosylated monomer of irisin (Figure 1). Through biophysical and biochemical experiments, refined by multiple steps of molecular dynamics simulations, Mu et al. were able to generate and refine a docking model with 2.98 Å root-mean-square deviation (RMSD) of the irisin/ $\alpha V\beta 5$ complex. This structure has impor-

tant implications for integrin-small ligand dynamics and how irisin mediates its physiological effects.⁵

Hsp90 molecular chaperones are important regulators of signal transduction pathways through their impact on the stabilization and activation of dependent proteins, termed “clients.”^{7,8} The model described by Mu et al. is reminiscent of previous studies on Hsp90 regulation of client protein ligand affinity. Steroid hormone receptors (SHRs) are a subset of Hsp90 client proteins, and interaction with Hsp90 chaperone complexes promotes ligand affinity for SHRs.⁹ Additionally, a previous report demonstrated that eHsp90, in complex with its dedicated extracellular co-chaperone TIMP2, enhanced binding to extracellular client MMP2 and regulated its proteinase activity.¹⁰ Mu et al. have demonstrated an analogous mechanism in which eHsp90 α 's interaction with $\alpha V\beta 5$ integrin promotes interaction with its ligand, irisin. These works collectively introduce a concept for Hsp90 in shaping substrates binding to their ligands.⁵

In summary, Mu et al. showed how a small polypeptide hormone, which exists in the extracellular space at much lower concentration compared to other matrix proteins (the canonical integrin ligands), uses integrin as the receptor to mediate downstream cellular events (Figure 1). This work describes a new two-step mechanism in which a hormone ligand acts through an integrin with the help of a differentially regulated third component.

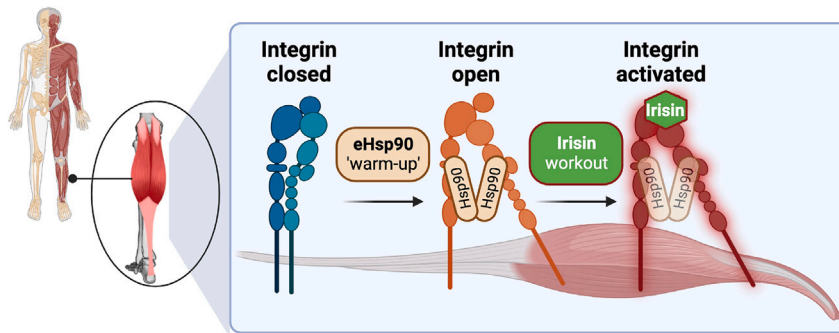


Figure 1. Extracellular chaperone Hsp90 “warms up” integrin for irisin workout

Physical exercise leads to an increase in irisin levels as well as release of the molecular chaperone Hsp90. Irisin alone has low affinity for the closed state of $\alpha V\beta 5$ integrin receptor. Extracellular Hsp90 (eHsp90) “opens” $\alpha V\beta 5$, allowing for irisin binding with high-affinity and effective signal transduction through its integrin receptor.

ACKNOWLEDGMENTS

This work was supported by the National Institute of General Medical Sciences of the National Institutes of Health under award numbers R01GM139932 (D.B.) and R35GM139584 (M.M.). The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

DECLARATION OF INTERESTS

The authors declare no competing financial interests.

REFERENCES

1. Baar, K., Wende, A.R., Jones, T.E., Marison, M., Nolte, L.A., Chen, M., Kelly, D.P., and Holloszy, J.O. (2002). Adaptations of skeletal muscle to exercise: rapid increase in the transcriptional

coactivator PGC-1. *FASEB J.* *16*, 1879–1886. <https://doi.org/10.1096/fj.02-0367.com>.

2. Boström, P., Wu, J., Jedrychowski, M.P., Korde, A., Ye, L., Lo, J.C., Rasbach, K.A., Boström, E.A., Choi, J.H., Long, J.Z., et al. (2012). A PGC1- α -dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature* *481*, 463–468. <https://doi.org/10.1038/nature10777>.
3. Islam, M.R., Valaris, S., Young, M.F., Haley, E.B., Luo, R., Bond, S.F., Mazuera, S., Kitchen, R.R., Caldaroni, B.J., Bettio, L.E.B., et al. (2021). Exercise hormone irisin is a critical regulator of cognitive function. *Nat. Metab.* *3*, 1058–1070. <https://doi.org/10.1038/s42255-021-00438-z>.
4. Kim, H., Wrann, C.D., Jedrychowski, M., Vidoni, S., Kitase, Y., Nagano, K., Zhou, C., Chou, J., Parkman, V.J.A., Novick, S.J., et al. (2018). Irisin mediates effects on bone and fat

via αV integrin receptors. *Cell* *175*, 1756–1768.e17. <https://doi.org/10.1016/j.cell.2018.10.025>.

5. Mu, A., Wales, T.E., Zhou, H., Draga-Coletă, S.V., Gorgulla, C., Blackmore, K.A., Mittenbühler, M.J., Kim, C.R., Bogoslavski, D., Zhang, Q., et al. (2023). Irisin acts through its integrin receptor in a two-step process involving extracellular Hsp90 α . *Mol. Cell* *83*, 1903–1920.e12. <https://doi.org/10.1016/j.molcel.2023.05.008>.
6. Di Virgilio, F., Sarti, A.C., Falzoni, S., De Marchi, E., and Adinolfi, E. (2018). Extracellular ATP and P2 purinergic signalling in the tumour microenvironment. *Nat. Rev. Cancer* *18*, 601–618. <https://doi.org/10.1038/s41568-018-0037-0>.
7. Schopf, F.H., Biebl, M.M., and Buchner, J. (2017). The HSP90 chaperone machinery. *Nat. Rev. Mol. Cell Biol.* *18*, 345–360. <https://doi.org/10.1038/nrm.2017.20>.
8. Dean, M.E., and Johnson, J.L. (2021). Human Hsp90 cochaperones: perspectives on tissue-specific expression and identification of cochaperones with similar in vivo functions. *Cell Stress Chaperones* *26*, 3–13. <https://doi.org/10.1007/s12192-020-01167-0>.
9. Backe, S.J., Sager, R.A., Regan, B.R., Sit, J., Major, L.A., Bratslavsky, G., Woodford, M.R., Bourboulia, D., and Mollapour, M. (2022). A specialized Hsp90 co-chaperone network regulates steroid hormone receptor response to ligand. *Cell Rep.* *40*, 111039. <https://doi.org/10.1016/j.celrep.2022.111039>.
10. Baker-Williams, A.J., Hashmi, F., Budzyński, M.A., Woodford, M.R., Gleicher, S., Himanen, S.V., Makedon, A.M., Friedman, D., Cortes, S., Namek, S., et al. (2019). Co-chaperones TIMP2 and AHA1 competitively regulate extracellular HSP90:client MMP2 activity and matrix proteolysis. *Cell Rep.* *28*, 1894–1906.e6. <https://doi.org/10.1016/j.celrep.2019.07.045>.